Louisiana Office of Public Health Laboratories		
Test Name	Rule out Anthrax	
PHL Location	Office of Public Health Laboratory Baton Rouge	
CPT Code	N/A	
Synonyms	BT Test, Bioterrorism Test, Clinical Rule Out	
Brief Description of Test	Prior notification requested. Contact Infectious Disease Epidemiology at 800-256-2748 or BT Program Advisor at 225-219-5241.  To rule out or confirm bacterial isolates as <i>Bacillus anthracis</i> or to rule out or confirm presence/absence of <i>Bacillus anthracis</i> in direct patient specimens.  B. anthracis is suspected and cannot be ruled out if the isolate fulfills the following characteristics:  • Direct Gram staining reveals large Gram-positive rods  • Grows on BAP as non-pigmented, odorless, white colonies having a ground glass appearance with edges that are slightly undulate ("Medusa heads"; comma shaped)  • Does not grow on MAC (or EMB)  • Non-hemolytic on BAP  • Catalase – positive  • Motility – negative	
Possible Results	Direct Sample Testing Presumptive Positive Equivocal Inconclusive Negative  Culture Isolate Testing Confirmed Positive Equivocal Negative	
Reference Range	Negative	
Specimen Type	Direct Sample – clinical specimens require prior approval	

	A. Collection and T	ransport of Clinical Specimens for Laboratory Rule-Out Testing*
	Cutaneous Vesicular (early) stage	Unroof vesicle and aspirate fluid or collect with two sterile swabs (dacron)
	Eschar (late) stage	<ul> <li>Insert swab (dacron) beneath the edge of the eschar, rotate swab or obtain an aspirate</li> <li>Transport specimens at room temperature</li> </ul>
	Gastrointestinal	Stool (> 5 gramspecan size), collect and transport in a leak proof sealed container     Collect blood (late stage of infection) directly into an appropriate blood culture bottle (aerobic and anaerobic)     Transport specimens and bottles at room temperature
	Inhalational	Sputum     Blood: collect directly into an appropriate blood culture bottle (aerobic and anaerobic)     Cerebral Spinal Fluid only if signs of meningitis occur     Transport specimens and bottles at room temperature
	Postmortem Tissue	<ul> <li>Tissue pieces should be collected and kept moist</li> <li>Transport in sterile container at room temperature within 1 hour of collection</li> </ul>
	Culture Isolate Actively growing	, pure culture inoculated in/on unexpired media
Specimen Container(s):	Specimen must b	e transported in "triple" packaging (primary receptacle, water ackaging and durable outer packaging) required for a biological
Minimum volume accepted:	1mL of blood, 0.2 a pea	Ig of tissue, 5mL of sputum or solids approximately the size of
	concentrations of B. anthracis can I nonclinical (envi reference laborate	5th Edition of the BMBL, unless you are working with high this organism or performing procedures that produce aerosols, be handled using BSL-2 practices. Do not process ronmental or animal) specimens in hospital or commercial pries; restrict processing to human clinical specimens only. mens should be directed to the designated LRN Reference
Collection Instructions	wearing gloves an (BSC) are recom- aerosol production be taped/shrink se	nens can be handled using BSL-2 practices. BSL-3 precautions, and gown and working in a certified Class II biosafety cabinet mended when performing activities having a high potential for $n$ . Subcultures should be performed in a BSC and plates should ealed, and incubated in $5-10\%$ CO2. All additional testing med only in the BSC while wearing gloves to prevent acquiring the skin.
	solution of 10% b microscopic slide	of laboratory surfaces is easily accomplished using a fresh bleach. In addition, pipettes, needles, plastic loops, and is should be soaked in 10% bleach or $10-30\%$ formalin for 24h claved. Phenolics are not sporocidal at the usual working
	For swab samples of cotton.	s, use nylon, polyester (e.g. Dacron), rayon or foam swabs instead

	<u> </u>
	Label specimen with Patient Name and a 2nd Unique Identifier such as a chart number or medical record number. DOB is not considered unique.
	Complete a LAB Form 93 to accompany the sample. Lab submission form must be thoroughly completed with patient's first and last name, 2 <sup>nd</sup> patient identifier, gender, date of birth, date and time of collection, specimen source, test requested, submitter's name, address, fax and contact number. Additional information regarding patients' address is requested.
	The same two unique identifiers <b>MUST</b> be recorded on the tube <b>AND</b> the Lab 93 form.
	Transport specimen to laboratory as soon as possible after collection/incubation. Keep submission forms insulated from specimens.
Storage and Transport Instructions	Culture may be shipped ambient. For storage and transport of clinical samples, contact <b>BT Program Advisor at 225-219-5241.</b>
	Send sample to the Office of Public Health Laboratory Baton Rouge, 1209 Leesville Avenue, Baton Rouge, LA 70802
	LRN (Laboratory Response Network) guidance for the packaging and shipping of infectious substances and biological agents should be consulted for recent changes. IATA and DOT publications continue to be revised frequently. Submitters should frequently and regularly consult IATA publications, the Federal Register, and the publications of other governing agencies for more complete instructions. It is the shipper's responsibility to ensure adherence to the most current regulations.
	Useful web sites that address the shipping of infectious substances and biological agents: International Air Transport Association: <a href="https://www.iata.org">https://www.iata.org</a> Department of Transportation: <a href="http://phmsa.dot.gov">http://phmsa.dot.gov</a> American Society for Microbiology: <a href="http://www.asm.org">http://www.asm.org</a> American Biological Safety Association: <a href="http://www.absa.org/">http://www.absa.org/</a> Animal and Plant Health Inspection Service: <a href="http://www.aphis.usda.gov/">http://www.aphis.usda.gov/</a> Centers for Disease Control and Prevention: <a href="http://www.cdc.gov/od/ohs/">http://www.cdc.gov/od/ohs/</a>
Causes for Rejection	Incorrect source     Incorrect labeling     Expired collection tubes     Not approved for testing by Infectious Disease Epidemiology
Limitations of the Procedure	If inhibitors are present in a DNA extraction, PCR assays may produce a false negative result.  A false negative result may occur if a sample is improperly collected, transported or handled. False negative results may occur if inadequate numbers of organisms are present in the sample.  Data suggest that clinical specimens collected subsequent to initiation of

	antimicrobial treatment may not be positive for <i>B. anthracis</i> due to reduction of <i>B. anthracis</i> organisms and DNA. Whenever possible, specimens collected prior to administration of antimicrobial agents should be used to determine infection with <i>B. anthracis</i> .  Real-time PCR results are considered presumptive unless a pure culture isolate exhibiting colony morphology and Gram stain characteristics (Gram positive rods) consistent with <i>B. anthracis</i> was used to generate the result.
Interfering Substances	N/A for direct culture testing
References	Sentinel Level Clinical Laboratory Guidelines for Suspected Agents of Bioterrorism and Emerging Infectious Diseases – <i>Bacillus anthracis</i> <a href="http://www.asm.org/images/PSAB/LRN/Anthrax316.pdf">http://www.asm.org/images/PSAB/LRN/Anthrax316.pdf</a> Laboratory Response Network
Additional Information	If <i>B. anthracis</i> is suspected, contact Infectious Disease Epidemiology at 1-800-256-2748 or BT Program Advisor at 225-219-5241.
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Warning: If you have printed a copy of this information please be advised that the Louisiana Office of Public Health Laboratories website and methods are updated on a regular basis. Please check the online version of this document to ensure you are relying on the most recent release.

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